



Reducing or Eliminating Polysorbate Induced Anaphylaxis and Unwanted Immunogenicity in Biotherapeutics.

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ABSTRACT

An increasing use of biotherapeutics across a growing spectrum of neoplastic, autoimmune, and inflammatory diseases has resulted in a corresponding increase in hypersensitivity reactions. The origins of anaphylaxis are often attributed to undefined intrinsic properties of the biotherapeutic protein itself, ignoring the broader potential negative contributions of functional excipients, in particular polyoxyethylene containing surfactants such as polysorbate 80 and polysorbate 20 (Tween 80 and Tween 20). These surfactants allow biotherapeutics to meet the stringent challenges of extended shelf-life, increased solubility, protein aggregation prevention, reduced administration volume, and satisfactory reconstitution properties in the case of lyophilized biotherapeutics. The potential negative impact of certain functional excipients on product performance characteristics such as anaphylaxis and immunogenicity is often overlooked. While regulatory authorities understandably focus heavily on comparable efficacy in evaluating biosimilars, similar efficacy does not necessarily imply a similar safety profile between the originator and biosimilar products. Both unwanted immunogenicity and anaphylaxis do comprise major components of safety assessment, however, few if any attempts are made to differentiate drug-related from excipient-related anaphylaxis. The replacement of anaphylactogenic and immunogenic functional excipients with equally effective but safer alternatives will allow biotherapeutic developers to differentiate their biotherapeutic, biosimilar, or biobetter from the large number of nearly identical competitor products, simultaneously providing a substantial commercial benefit as well as critical clinical benefits for all concerned, that is, patients, physicians, and third party payers.

KEY WORDS: Polysorbate, Tween-80, functional excipient, allergic reaction, anaphylaxis, hypersensitivity, immunogenicity, alkylsaccharide, polyoxyethylene, surfactant

INTRODUCTION

Biosimilars are important contributors to improved healthcare and as more and more are receiving regulatory approval, they are gaining

greater acceptance and broader utility. The rapidly increasing use of monoclonal antibodies in the treatment of neoplastic, autoimmune, and inflammatory diseases has resulted in a dramatic increase in hypersensitivity reactions worldwide, complicating the use of first-line therapies and impacting patients' survival and quality of life (1).

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The origins of anaphylaxis are not well understood and are often attributed to undefined intrinsic properties of the biotherapeutic, in spite of the fact that biotherapeutic formulations are necessarily complex, employing a host of functional excipients beyond the biotherapeutic protein itself. These functional excipients are necessary in order to meet the stringent challenges of extended shelf-life, stabilization, solubility, aggregation prevention, especially at the high concentrations typically used to reduce administration volume and time, and in the case of lyophilized products, satisfactory reconstitution properties. The potential negative impact of certain functional excipients on product performance characteristics such as anaphylaxis and immunogenicity is often overlooked. For example, a title search on PubMed, The U.S. National Library of Medicine, using the truncated stem biotherapeutic* in combination with the truncated stems anaphyla* or hypersensit* or allerg* yielded 332 journal articles with biotherapeutic/biotherapeutics in the title, but only a single combination with any of the other stems. Similarly, a title search using the truncated stem biosimilar* in combination with the truncated stems anaphyla* or hypersensit* or allerg* yielded 1218 journal articles having biosimilar/biosimilars in the title, and only a single return was received, in this case relating to anaphylaxis of Glargine, a modified human insulin. Interestingly, Glargine contains a polysorbate surfactant. Two other commonly prescribed human insulins, Humalog and Humulin, do not contain polysorbate surfactants and do not show any reports of anaphylaxis. The NLM MedLine searches were conducted between May 11 and May 15, 2017 and reflect NLM database content within this time frame.

To examine the level of interest in the role of polysorbates, the most common class of surfactant excipients used in formulating biotherapeutics, in connection with anaphylaxis, a title search using the truncated stem

polysorbate* in combination with the truncated stems anaphyla* or hypersensit* or allerg* or immunogenic* yielded 362 journal articles with polysorbate/polysorbates in the title, with 4, 4, 1, and 1 in combination with anaphyla* or hypersensit* or allerg*, respectively (i.e., 10 articles returned in total with 9 being unique). Of this small number of articles dealing with anaphylaxis, 6 reported observations of human anaphylactic/allergic responses and three related to *in vivo* animal studies.

Polysorbates are used in formulations of non-biotherapeutic drugs as well. Norris *et al.* conducted extensive analysis of the rates of anaphylaxis for the taxanes docetaxel and paclitaxel (2). Docetaxel is formulated with PS-80 and paclitaxel is formulated with Cremophor EL, another polyoxyethylene based surfactant that is not typically used in biotherapeutic formulations but often used in small molecule drug formulations. The taxanes are small molecule drugs, not biotherapeutics, however, like many biotherapeutics, hypersensitivity reactions are common side effects. Because the hypersensitivity reactions do not appear to be IgE mediated, it is hypothesized that the polyoxyethylene-based surfactants are the causative agents.

The reported rate of hypersensitivity reactions with docetaxel is estimated at 30% for patients who do not receive pre-medications such as dexamethasone. Of 67 cases of docetaxel-induced anaphylaxis, 34% represented fatalities. Paclitaxel induced hypersensitivity reactions occur in up to 41% of patients. A more recently introduced taxane, cabazitaxel, also formulated with PS-80, lists severe hypersensitivity in its associated the Black Box warning, and it is suggested that an antihistamine, a corticosteroid, and an H₂ blocker be administered prophylactically.

The development cost of a biosimilar is not insignificant and is expected to represent a much larger fraction of the corresponding development cost and ultimate pricing of small

molecule generics. While this has been cited as a negative aspect of biosimilars, it opens the door to another entirely different perspective. Since the development cost in some cases may already be relatively high, the additional cost of modifying the formulation to improve performance, for example by circumventing the use of anaphylactogenic or immunogenic functional excipients, represents a proportionately smaller fraction of the total development cost. And in cases where high development costs will make price competition a less desirable commercial strategy, product acceptance, which ultimately translates to market share, will largely be driven by improved product performance characteristics rather than lower pricing.

The large, and growing number, of nearly identical biosimilars and 'biobetters', illustrated in Table 1 (3), provides a significant, if not essential, commercial incentive to differentiate one product from another. It should be noted that the term "biobetter" is not an approved term from a regulatory perspective. No separate regulatory pathway exists for 'biobetters' and they are treated essentially as New Chemical Entities. Nevertheless, the term is being used with increasing frequency. A simple intuitive definition is that 'biobetters' are improvements to originator biological molecules, offering some degree of superiority over the originator drug and its biosimilar competitors (4). Anour provides an illustrative example of a biobetter in the form of Roche's anti-CD20 monoclonal antibody obinutuzumab, which has shown superior efficacy in the treatment of chronic lymphocytic leukemia (CLL) compared to its 'originator' rituximab (5). Barbosa *et al.* reviewed the process for the development of biosimilars and 'biobetters', focusing on how the various steps impact immunogenicity and the development of anti-drug antibodies (6). While the total market for many of these originator products is substantial, the likelihood that more than a small handful of each competitive biosimilar product will achieve a

Table 1 Some major recombinant originator products and classes with biosimilars and biobetters currently in development

PRODUCT OR PRODUCT CLASS	SALES (US\$,B)	BIOSIMILARS	BIOBETTERS
Humira	\$9.27	13	7
Remicade	\$8.90	9	9
Enbrel	\$7.87	21	8
Rituxin	\$7.29	30	17
Herceptin	\$6.40	24	12
Lantus	\$6.40	5	2
Avastin	\$6.26	14	9
Neulastin	\$4.10	14	9
Lucentis	\$3.73	2	2
Aransap	\$3.00	4	2
Epogen	\$3.73	69	26
Novoseven	\$1.50	8	12
Neupogen	\$1.44	52	17
Insulin and analogs		40	53
Tumor necrosis factor (mAbs/inhibitors)		44	19
Interferons (alpha)		55	48
Interferons (beta)		23	23
Somatropins		28	17
Factors VIII		4	21
mAbs and antibody fragments		145	91
Cancer-targeted proteins (non-mAb)		264	159
Cancer-targeted mAbs		77	59

Source: Rader RA, An Analysis of the US Biosimilars Development Pipeline and Likely Market Evolution, Bio Process International, 11(6)s, June 2013, 16-23

significant percentage of the total market is slim. 'Biobetters' are likely to fare better than biosimilars if a clear benefit can be demonstrated in clinical trials. Minimal, or somewhat esoteric changes, for example related to longer shelf life or the elimination of the need to reconstitute in the pharmacy, are not likely to be persuasive in having a meaningful impact on the purchasing decisions of treating-physicians or institutional purchasers. In contrast, significant improvements, such as reduced or eliminated anaphylaxis and immunogenicity, or reduced likelihood of developing neutralizing antibodies in comparison to the originator biotherapeutic or

alternative biosimilars would likely have a very substantial and beneficial effect on prescribing and purchasing decisions.

Polysorbates are among the most common functional excipients in biotherapeutics

Surfactants, such as the polysorbates Tween-80 and Tween-20 (PS-80 and PS-20 respectively) are routinely used to prevent or limit the highly significant and very common problem of protein aggregation (7-19). Polysorbates are highly effective in preventing aggregation but polysorbates suffer from significant limitations. Specifically, polysorbate surfactants are not single or discrete chemical entities. They are complex and highly variable mixtures of fatty acid esters of polyoxyethylene sorbitan with residual amounts of polyoxyethylene sorbitan, polyoxyethylene, and isosorbide polyoxyethylene fatty acid esters, as well as, spontaneously forming protein-damaging hydro- and alkyl-peroxides, epoxy acids, and reactive aldehydes (20-25). Lot-to-lot variability in the concentration of chemically reactive species such as peroxides has been found to exceed an order of magnitude (26).

PS-80 is more prone to the generation of oxidative species compared with PS-20 as a result of the greater content of unsaturated alkyl side chains in PS-80 (27). In aqueous formulations polysorbates can also hydrolyze with an apparent half-life of five months at 40°C (28-30). While aggregation alone is known to cause unwanted immunogenicity, progressive oxidative damage to the protein's aminoacyl side chains caused by the contaminating peroxides, other reactive species cause neoantigen formation over time further increasing or altering immunogenicity. The peroxides principally damage methionine and tryptophan side chains (31-37). The aldehydes react with primary amino groups on proteins and are known to induce immunogenicity of proteins in the absence of aggregation or adjuvants. The epoxy acids can react with any exposed nucleophile. Oxidative damage also

leads to peptide bond hydrolysis, altering protein tertiary structure, yet another source of protein aggregation and releasing free fatty acids which can cause increased turbidity, a separate but not insignificant issue altogether (28,30).

Polysorbate induced anaphylaxis

Polysorbates, as well as certain polysorbate degradation products, are intrinsically anaphylactogenic. Examples of hypersensitivity reactions to polysorbates in humans and animal models, have been demonstrated by such indicators as histamine release, hemodynamic effects, skin prick testing, enzyme-linked immunosorbent assay, IgE immunoblotting, flow cytometric detection of basophil activation, complement activation, determination certain humoral factors, and the absence of polysorbate specific IgE (to confirm the non-immunologic nature of the anaphylactoid reactions). These are summarized below.

Anaphylaxis is a serious allergic reaction associated with the administration of some biotherapeutics. There has been minimal research into the specific causes in the case of either the originator or biosimilar biotherapeutics, as indicated above by the small number of articles dealing with the subject. One may reasonably speculate that since proteins are well known initiators of anaphylaxis, the role of other formulation components, such as surfactants in initiating anaphylaxis has generally been overlooked. While polysorbate-induced unwanted immunogenicity is well documented in the literature (30, 38-40) little attention has been paid to polysorbate induced anaphylaxis.

Anaphylaxis symptoms occur over minutes to hours with an average onset of 5 to 30 minutes if exposure is intravenous, as it is for many biotherapeutics, and can affect the skin, respiratory system, gastrointestinal tract, the heart and vasculature, and the central nervous

system. Symptoms include hives, itching, flushing, or swelling (angioedema), swelling of the tongue or throat, runny nose and swelling of the conjunctiva. Respiratory symptoms include shortness of breath, bronchial spasm, and upper airway obstruction (41-47). Coronary artery spasm and associated drop in blood pressure or shock, and subsequent myocardial infarction, dysrhythmia, or cardiac arrest may also occur. In some cases it may cause death (44,45).

Intramuscularly injected epinephrine is the primary treatment for anaphylaxis along with antihistamines, steroids, intravenous fluids, and positioning the person flat (41,46). While regulatory authorities understandably focus heavily on comparable efficacy in evaluating biosimilars, similar efficacy does not necessarily imply a similar safety profile between the originator and biosimilar products. Both unwanted immunogenicity and anaphylaxis do comprise major components of the safety assessment requirements of biotherapeutics, however, few or no attempts have been made to distinguish functional excipient-induced anaphylaxis from anaphylaxis arising from the biotherapeutic protein itself. In the case of monoclonal antibodies, drug hypersensitivity and anaphylaxis have been reported for rituximab, ofatumumab, obinutuzumab, trastuzumab, cetuximab, tocilizumab, infliximab, etanercept, adalimumab, abciximab, golimumab, certolizumab, brentuximab, bevacizumab, and omalizumab, all of which contain a polysorbate surfactant (1).

PS-80 and PS-20, found in more than 70% of monoclonal antibody and other protein biotherapeutic drugs, have now been shown to cause anaphylaxis in patients receiving biotherapeutics. The anaphylactogenic properties of PS-80 are now increasingly well documented in the clinical literature. The precise mechanistic cause, or causes, of polysorbate induced anaphylaxis is complicated by their complex chemical nature. A number of specific molecular species that induce

anaphylaxis have been identified in preclinical animal studies. For example, as early as 1985, Masini *et al.* demonstrated that polysorbate induced histamine release in peripheral tissues and isolated mast cells, as well as hemodynamic responses (47). In 1997, Bergh *et al.* reported that air exposure of aqueous solutions of PS-80 produced formaldehyde and acetaldehyde (48), the latter shown to react with proteins and produce anaphylactogenic moieties (49) and be anaphylactogenic in amounts that may be eliciting allergic reactions in some individuals. The authors prophetically warned that allergenic compounds are formed during manufacture, storage, and handling of products containing polysorbates and chemically similar surfactants and that this should be taken into consideration by drug developers.

Coors *et al.* conducted a thorough examination of PS-80 as an inducer of severe anaphylaxis in patients receiving intravenous drug formulations. They employed an extensive complement of well accepted and sensitive detection methodologies including skin prick testing, enzyme-linked immunosorbent assay, IgE immunoblotting, and flow cytometric detection of basophil activation, in control patients and patients with a medical history of anaphylactic shock due to intravenous administration of a multivitamin product as a surrogate for intravenously administered drugs. Polysorbate specific IgE antibodies were not identified in enzyme-linked immunosorbent assay and immunoblot examinations, confirming the nonimmunologic nature of the anaphylactoid reaction. Their study unambiguously demonstrated that PS-80 can cause severe nonimmunologic anaphylactoid reactions (50).

Sun *et al.* evaluated the sensitization effect of PS-80 from different manufacturing lots in beagle dogs. Varying degrees of anaphylactoid reaction were observed (51). Similarly, Yang *et al.* assessed 10 batches of PS-80 solutions from various suppliers and found that spontaneously formed PS-80 impurities such as peroxides and

oxidized fatty acid residues, present in varying levels in each of the tested batches, induced anaphylactoid reactions in an *in vivo* zebra fish model (52).

Qiu *et al.* demonstrated that polysorbate 80 induces typical non-immune anaphylactic reactions (pseudoallergy) in dogs characterized by the release of histamine and unvaried IgE antibodies. PS-80 induced the release of histamine, a 2-fold increase in SC5b-9, a 2.5-fold increase in C4d, and a 1.3-fold increase in Bb, while IgE remained unchanged. PS-80 caused cardiopulmonary distress in dogs and activated the complement system through classical and alternative pathways as indicated in both *in vivo* and *in vitro* assays (53).

With the increasing importance and routine use of a growing number of biotherapeutics, clinical reports describing polysorbate induced anaphylaxis are increasing as well. For example, in two patients receiving omalizumab, reactions after administration have been reported. Intradermal testing produced significant wheal/flare reaction to PS-20 but not in the negative control subject. The *in vitro* and *in vivo* immunologic data support the conclusion that the adverse reactions experienced by the two patients after more than a year of successful omalizumab therapy were likely anaphylactoid in nature (54). An earlier report of unexplained omalizumab anaphylaxis appeared but the possible association with PS-20 was not considered at that time (55).

Patients receiving the red cell growth hormones darbepoietin and erythropoietin developed hypersensitivity reactions. Based on subsequent skin testing and the observed clinical effects it was concluded that the cause of these reactions was due to the excipient PS-80, and that this might have contributed to the incidence of pure red cell aplasia (56).

In a study comparing etoposide formulations with and without PS-80 using the same premedication protocol, the patient exhibited

hypersensitivity reaction with the PS-80 containing formulation but none with the etoposide formulation not containing PS-80. The authors concluded that the hypersensitivity reaction was likely due to PS-80 rather than the etoposide itself (57).

Badiu *et al.* reported multiple cases of PS-80 induced anaphylaxis arising from administration of different vaccines. A female patient experienced generalized urticaria, eyelid angioedema, rhino-conjunctivitis, dyspnoea and wheezing 1 hour after her third intramuscular dose of quadrivalent human papilloma virus vaccine, Gardasil, which contains PS-80 (58). Gardasil also yielded positive intradermal tests, while skin tests with the bivalent vaccine not containing PS-80 were negative. Prick tests performed with PS-80 were positive in the patient and negative in ten healthy controls. The CD203 basophil activation test result was negative for PS-80 at all the tested dilutions and specific IgE was not found. The authors also skin tested patients receiving two flu vaccines, one containing PS-80 (Fluarix, GSK), which resulted in a positive reaction and another flu vaccine with no adjuvant or preservative (Vaxigrip, Sanofi Pasteur MSD), which yielded negative results.

Limaye *et al.* reported an allergic reaction to erythropoietin which included generalized pruritis, erythema, and orofacial angioedema (59). The Eprex erythropoietin formulation contained recombinant human erythropoietin and PS-80 as excipient (0.15 mg/ml). Skin prick and sequential intradermal testing with increasing concentrations of Eprex and Neupogen (Amgen, Thousand Oaks, CA) also containing polysorbate at 0.04 mg/ml, gave positive reactions, whereas a polysorbate-free erythropoietin preparation yielded negative test results. Intradermal testing with pharmaceutical-grade polysorbate resulted in a positive local reaction followed by mild orofacial angioedema 1 hour later. No reaction was observed in a control subject. Purcell *et al.* identified polysorbate 80 as the likely cause of

immune response to erythropoietin when human albumin was replaced by polysorbate 80 and glycine (60). In addition to the vaccine examples cited above, polysorbate induced anaphylactic responses have also been reported in non-biologic drug classes containing polysorbate, such as vitamin A, certain steroids, and acyclovir (61, 62).

Separating mab induced anaphylaxis from PS induced anaphylaxis

While it is clear that polysorbates can and do induce anaphylactic responses, current clinical trials do not appear to attempt to differentiate polysorbate-induced anaphylaxis from biotherapeutic-induced anaphylaxis. In order to do this, separate vehicle studies would have to be conducted. Considering the fact that anaphylaxis only occurs in a fraction of patients, and patients as a whole will likely have had varying exposure histories to polysorbates for earlier, and perhaps unrelated, disease treatments, selecting multiple control cohorts, i.e., no previous polysorbate exposure *versus* previous polysorbate exposure over varying timeframes, is difficult and prohibitively expensive and would add further costs to the already expensive clinical trial and regulatory approval process. Substituting alternative stable, non-chemically reactive, non-anaphylactogenic surfactants such as alkylsaccharides (38, 40, 63, 64) in biotherapeutics, and comparing anaphylaxis rates in clinical trials would begin to answer this question.

CONCLUSION

Replacing polysorbates with surfactants that minimize anaphylaxis episodes, and at the same time, do not result in progressive protein degradation and increased immunogenicity, would meet a critical need while providing a substantial differentiating clinical benefit for all concerned, that is, patients, physicians, and third party payers. The most advanced alternative appears to be a class of non-ionic surfactants termed alkylsaccharides.

Alkylsaccharides are comprised of a sugar coupled to an alkyl chain (35, 37, 59, 60). While certain alkylsaccharides are GRAS for use in food and cosmetic products, no biotherapeutics employing alkylsaccharides have yet been approved. Impediments to approval of any functional excipient, including potential alternative surfactants, include the significant costs of demonstrating functional effectiveness, safety and tolerability, as well as the cost of establishing and qualifying multiple GMP manufacturing sources in order to guarantee an uninterrupted supply during manufacturing.

'Biobetters' may offer the benefit of earlier market entry compared to the corresponding biosimilar because the product launchability date for a biosimilar is determined by the latest relevant patent expiration date as well as any data and market exclusivity extensions. Because 'biobetters' involve full approvals, they are not affected by data exclusivity, so that their launchability dates would be determined solely by the latest date of patent expiration or market exclusivity grants (3).

The clinical and commercial incentives to replace polysorbates are clear. For originator biotherapeutics, minimization of anaphylaxis would offer significant clinical and safety benefits to patients, possibly reducing the time and cost of pretreatment with antihistamines and steroids. In the case of biosimilars, risks associated with high development cost, continued pricing uncertainty, with pricing estimates of ranging from 25% to 75% of the originator product price (65), and the large number of competitor products, with over 900 biosimilars and 600 'biobetters' currently in development, the ability to differentiate a product based upon improved clinical or safety characteristics, may be critical to commercial success. Commercial success is key to achieving the objectives of reduced biotherapeutic cost and increased patient accessibility. The ability to offer patients some clear and substantial clinical benefit would likely facilitate the physician's decision or recommendation to switch a patient

away from a well characterized originator product to a newly introduced 'biobetter'.

CONFLICT OF INTEREST STATEMENT

Dr. Maggio is the CEO of Aegis Therapeutics LLC (Aegis), as well as, a shareholder, and serves on its Board of Managers. Aegis commercializes drug formulation technologies based upon the use of alkylglycosides which were developed at the University of Alabama Medical Center, Birmingham, Alabama, by Professors Dennis Pillion and Eli Meezan. Issued Aegis patents cover the use of alkylsaccharides as absorption enhancers for peptide and protein drugs via intranasal and oral routes, and the prevention of aggregation of protein and peptide-based drugs, including monoclonal antibodies. Aegis is a party to a number of research and commercialization licenses with multinational pharmaceutical companies active in the biotherapeutics field.

REFERENCES

- Bonamichi-Santos R, Castells M. Diagnoses and Management of Drug Hypersensitivity and Anaphylaxis in Cancer and Chronic Inflammatory Diseases: Reactions to Taxanes and Monoclonal Antibodies." *Clin Rev Allergy Immunol*, doi:10.1007/s12016-016-8556-5, 2016.
- Norris LB, Qureshi ZP, Bookstaver PB, Raisch DW, Sartor O, Chen H, Chen F, Bennett CL. Polysorbate 80 hypersensitivity reactions: a renewed call to action. *Community Oncology*, 7(9): 425-428, 2010.
- Rader RA. An analysis of the US Biosimilars Pipeline and Likely Market Evolution. *BioProcess International*. 11(6)s: 16-23, 2013.
- Biobetters rather than biosimilars. Posted 06/05/2011. <http://gabi-journal.net/biosimilars-versus-biobetters-a-regulators-perspective.html>
- Anour R. Biosimilars versus 'biobetters'—a regulator's perspective. *Generics and Biosimilars Initiative Journal, GaBI Journal*, 3(4):166-7, 2014.
- Barbosa MD, Kumar S, Loughrey H, Singh SK. Biosimilars and biobetters as tools for understanding and mitigating the immunogenicity of biotherapeutics. *Drug Discov Today*, 17(23-24):1282-8, 2012.
- Roy S, Jung R, Kerwin BA, Randolph TW, Carpenter JF. Effects of Benzyl Alcohol on Aggregation of Recombinant Human Interleukin-1-Receptor Antagonist in Reconstituted Lyophilized Formulations. *J Pharm Sci*, 94(2):382–396, 2005.
- Clodfelter DK, Pekar AH, Rebhun DM, Destrampe KA, Havel HA, Myers SR, Brader ML. Effects of Non-Covalent Self-Association on the Subcutaneous Absorption of a Therapeutic Peptide. *Pharm Res*, 15(2): 254–262, 1998.
- DePalma A. BioProcessing: Improving Stability While Adding Value. *Gen Eng News*, 15 January 2006.
- Casadevall N, Nataf J, Viron B, Kolta A, Kiladjian JJ, Martin-Dupont P, Michaud P, Papo T, Ugo V, Teyssandier I, Varet B, Mayeux P. Pure Red-Cell Aplasia and Antierythropoietin Antibodies in Patients Treated with Recombinant Erythropoietin. *NEJM* 346(7): 469–475, 2002.
- Hermeling S, Schellekens H, Maas C, Gebbink MF, Crommelin DJ, Jiskoot W. Antibody Response to Aggregated Human Interferon Alpha2b in Wild-Type and Transgenic Immune Tolerant Mice Depends on Type and Level of Aggregation. *J Pharm Sci*, 95(5): 1084–1096, 2006.
- Vartanian T, Sorensen PS, Rice G. Impact of Neutralizing Antibodies on the Clinical Efficacy of Interferon Beta in Multiple Sclerosis. *J Neurology*, 251: Suppl. II 25–30, 2004.
- Pezron I, Mitra R, Pal D, Mitra AK. Insulin Aggregation and Asymmetric Transport Across Human Bronchial Epithelial Cell Monolayers (Calu-3). *J Pharm Sci*, 91(4): 1135–1146, 2002.
- Sluzky V, Tamada JA, Klivanov AM, Langer R. Kinetics of Insulin Aggregation in Aqueous Solutions Upon Agitation in the Presence of Hydrophobic Surfaces. *PNAS USA*, 88(21): 9377–9781, 1991.
- King HD, Dubowchik GM, Mastalerz H, Willner D, Hofstead SJ, Firestone RA, Lasch SJ, Trail PA. Monoclonal Antibody Conjugates of Doxorubicin Prepared with Branched Peptide Linkers: Inhibition of Aggregation By Methoxytriethelene glycol Chains. *J Med Chem*, 45(19): 4336–4343, 2002.
- Konarkowska B, Aitken JF, Kistler J, Zhang S, Cooper GJ. The Aggregation Potential of Human Amylin Determines Its Cytotoxicity Towards Islet Beta-Cells. *Febs J*, 273: 3614–3624, 2006.
- Elgersma RC, Meijneke T, Posthuma G, Rijkers DT, Liskamp RM. Self-Assembly of Amylin(20–29) Amide-Bond Derivatives into Helical Ribbons and Peptide Nanotubes Rather Than Fibrils. *Chemistry*. 2006; 12: 3714–3725, 2006.
- Purohit VS, Middaugh CR, Balasubramanian SV. Influence of Aggregation on Immunogenicity of

- Recombinant Human Factor VIII in Hemophilia A Mice. *J Pharm Sci*, 95(2): 358–371, 2006.
- 19 Porter WR, Staack H, Brandt K, Manning MC. Thermal Stability of Low Molecular Weight Urokinase During Heat Treatment: I. Effects of Protein Concentration, pH and Ionic Strength. *Thromb Res*. 71(4): 265-79, 1993.
 - 20 Hawe A, Filipe V, Jiskoot W. Fluorescent Molecular Rotors as Dyes to Characterize Polysorbate-Containing IgG Formulations. *Pharm Res*, 27(2): 314-326, 2010.
 - 21 Kerwin BA. Polysorbates 20 and 80 Used in the Formulation of Protein Biotherapeutics: Structure and Degradation Pathways. *J Pharm Sci*, 97: 2924–2935, 2008.
 - 22 Ayorinde FO, Gelain SV, Johnson JH Jr, Wan LW. Analysis of Some Commercial Polysorbate Formulations Using Matrix Assisted Laser Desorption/Ionization Time-of-Flight Mass Spectrometry. *Rapid Commun Mass Spectrom*, 14: 2116–2124, 2000.
 - 23 Brandner JD. The Composition of NF-Defined Emulsifiers: Sorbitan Monolaurate, Monopalmitate, Monostearate, Monooleate, Polysorbate 20, Polysorbate 40, Polysorbate 60, and Polysorbate 80. *Drug Dev Ind Pharm*, 24: 1049–1054, 1998.
 - 24 Frison-Norrie S, Sporns P. Investigating the Molecular Heterogeneity of Polysorbate Emulsifiers By MALDI-TOF MS. *J Agric Food Chem*, 49: 3335–3340, 2001.
 - 25 Zhang R, Wang Y, Tan L, Zhang HY, Yang M. Analysis of Polysorbate 80 and Its Related Compounds By RP-HPLC with ELSD and MS Detection. *J Chromatogr Sci*, 50(7): 598–607, 2012.
 - 26 Wasylaschuk WR, Harmon PA, Wagner G, Harman AB, Templeton AC, Xu H, Reed RA. Evaluation of Hydroperoxides in Common Pharmaceutical Excipients. *J Pharm Sci*, 96 (1): 106 – 116, 2007.
 - 27 Yao J, Dokuru DK, Noestheden M, Park SS, Kerwin BA, Jona J, Ostovic D, Reid DL. A quantitative kinetic study of polysorbate autooxidation: the role of unsaturated fatty acid substituents. *Pharm Res*, 6(10): 2303-2313, 2009.
 - 28 Donbrow M, Azaz E, Pillersdorf A. Autoxidation of polysorbates, *J Pharm Sci*, 67(12): 1676–1681, 1978.
 - 29 Chafetz L., Hong W-H., Tsilifonis D.C., Taylor A.K., Philip J., Decrease in the rate of capsule dissolution due to formaldehyde from Polysorbate 80 autoxidation. *J Pharm Sci*, 73: 1186-1187, 1984.
 - 30 Kishore R.S., Kiese S., Fischer S., Pappenberger A., Grauschopf U., Mahler H.C., The degradation of polysorbates 20 and 80 and its potential impact on the stability of biotherapeutics. *Pharm Res*, 28(5): 1194-210, 2011.
 - 31 Ji JA, Zhang B, Cheng W, Wang YJ. Methionine, Tryptophan, and Histidine Oxidation in a Model Protein, PTH: Mechanisms and Stabilization. *J Pharm Sci*. 98: 4485–4500, 2009.
 - 32 Simat TJ, Steinhart H. Oxidation of Free Tryptophan and Tryptophan Residues in Peptides and Proteins. *J Agric Food Chem*, 46: 490–498, 1998.
 - 33 Thiele GM, Tuma DJ, Willis MS, Miller JA, McDonald TL, Sorrell MF, Klassen LW. Soluble Proteins Modified with Acetaldehyde and Malondialdehyde Are Immunogenic in the Absence of Adjuvant. *Alcohol Clin Exp Res*, 22: 1731–1739, 1998.
 - 34 Moghaddam AE, Gartlan KH, Kong L, Sattentau QJ. Reactive Carbonyls Are a Major Th2-Inducing Damage-Associated Molecular Pattern Generated By Oxidative Stress. *J Immunol*, 187: 1626–1633 2011.
 - 35 Allison ME, Fearon DT. Enhanced Immunogenicity of Aldehyde-Bearing Antigens: A Possible Link Between Innate and Adaptive Immunity. *Eur J Immunol*, 30: 2881–2887, 2000.
 - 36 J.W. Chu, J. Yin, D.I.C. Wang, and B.L. Trout L. Understanding Oxidative Instability of Protein Pharmaceuticals. *Mol Eng Biol Chem Systems*, <http://hdl.handle.net/1721.1/3955>, 2004.
 - 37 Davies KJ. Protein Damage and degradation by oxygen radicals. *J Biol Chem*, 262: 9895-901.
 - 38 Maggio ET. Biosimilars, Oxidative Damage, and Unwanted Immunogenicity - A Review, *BioProcess International*, 11(6)s: 28-14, 2013.
 - 39 Ha E, Wang W, Wang YJ. Peroxide formation in polysorbate 80 and protein stability. *J Pharm Sci*, 91(10): 2252–2264, 2002.
 - 40 Maggio ET. Alkyl Mono- and Diglucosides Highly Effective, Nonionic Surfactant Replacements for Polysorbates in Biotherapeutics — A Review. *BioProcess International*, 14(3): 30-49, 2016.
 - 41 Simons FE. Anaphylaxis: Recent advances in assessment and treatment. *J Allergy and Clinical Immunology*. 124(4): 625–36, 2009.
 - 42 Oswalt ML, Kemp SF. Anaphylaxis: office management and prevention. *Immunol. Allergy Clin. North Am*, 27(2): 177–91, 2007.
 - 43 Brown, SG et al. Anaphylaxis: diagnosis and management. *The Medical Journal of Australia*, 185(5): 283–9, 2006.
 - 44 Lee JK, Vadas P. Anaphylaxis: mechanisms and management. *Clin Exp Allergy*, 42(7): 923-938, 2011.

- 45 Khan BQ, Kemp FS. Pathophysiology of Anaphylaxis. *Current Opinion in Allergy and Clinical Immunology*, 11(4): 319–25, 2011.
- 46 Simons, FE. World Allergy Organization survey on global availability of essentials for the assessment and management of anaphylaxis by allergy- immunology specialists in health care settings. *Annals of Allergy, Asthma & Immunology*, 104(5): 405–12, 2010.
- 47 Masini E, Planchenault J, Pezziardi F, Gautier P, Gagnol JP. Histamine-releasing properties of Polysorbate 80 in vitro and in vivo: correlation with its hypotensive action in the dog. *Agents Actions*, 16(6): 470-7, 1985.
- 48 Bergh M, Magnusson K, Nilsson JL, Karlberg AT. Contact allergenic activity of Tween 80 before and after air exposure. *Contact Dermatitis*, 37(1):9-18 1997.
- 49 Israel Y, MacDonald A, Niemelä O, Zamel D, Shami E, Zywlko M, Klajner F, Borgono C. Hypersensitivity to acetaldehyde-protein adducts. *Molecular Pharmacology*, 42 (4): 711-717, 1992.
- 50 Coors EA, Seybold H, Merk HF, Mahler V. Polysorbate 80 in medical products and nonimmunologic anaphylactoid reactions. *Ann Allergy Asthma Immunol*, 95(6):593-9, 2005.
- 51 Sun W, Li Y, Wang N, Du F, Hao W, Zhao L. Anaphylactoid reactions induced by polysorbate 80 on Beagle dogs. *Zhongguo Zhong Yao Za Zhi*), 36(14): 1874-1878, 2011.
- 52 Yang R, Lao QC, Yu HP, Zhang Y, Liu HC, Luan L, Sun HM, Li CQ. Tween-80 and impurity induce anaphylactoid reaction in zebrafish. *J Appl Toxicol*, 5(3): 295-301, 2015.
- 53 Qiu S, Liu Z, Hou L, Li Y, Wang J, Wang H, Du W, Wang W, Qin Y, Liu Z. Complement activation associated with polysorbate 80 in beagle dogs. *Int Immunopharmacol*, 15(1): 144-149, 2013.
- 54 Price KS, Hamilton RG. Anaphylactoid reactions in two patients after omalizumab administration after successful long-term therapy. Anaphylactoid reactions in two patients after omalizumab administration after successful long-term therapy. *Allergy Asthma Proc*, 28(3): 313-9, 2007.
- 55 Dreyfus DH, Randolph CC. Characterization of an anaphylactoid reaction to omalizumab. *Ann. Allergy Asthma Immunol*, 96(4): 624-7, 2006.
- 56 Steele RH, Limaye S, Cleland B, Chow J, Suranyi MG. Hypersensitivity reactions to the polysorbate contained in recombinant erythropoietin and darbepoietin. *Nephrology (Carlton)*, 10(3): 317-20, 2005.
- 57 Aksahin A, Colak D, Altinbaset M. Etoposide? Or polysorbate-80? *Indian J Cancer*, 48(2): 272-273, 2011.
- 58 Badiu I, Geuna M, Heffler E, Rolla G. Hypersensitivity reaction to human papillomavirus vaccine due to polysorbate 80. *BMJ Case Reports*, doi:10.1136/bcr.02.2012.5797, 2012.
- 59 Limaye S, Steele RH, Quin J, Cleland B. An allergic reaction to erythropoietin secondary to polysorbate hypersensitivity. *J Allergy Clin Immunol*, 110(3): 530, 2002.
- 60 Purcell RT, Lockley RF. Immunologic responses to therapeutic biologic agents. *J Investig Allergol Clin Immunol*, 18: 335–42, 2008.
- 61 Shelley WB, Talanin N, Shelley ED. Polysorbate 80 hypersensitivity. *Lancet*, 345(8960): 1312-1313, 1995.
- 62 Palacios Castano, MI, Venturini Díaz, M. Anaphylaxis Due to the Excipient Polysorbate 80. *J Investig Allergol Clin Immunol*, 26(6):394-396, 2016.
- 63 Maggio ET. Polysorbates, peroxides, protein aggregation, and immunogenicity – a growing concern. *J Excipients and Food Chem*. 2012;3(2):45-53, 2012.
- 64 Maggio ET. Alkylsaccharides: circumventing oxidative damage to biotherapeutics caused by polyoxyethylene-based surfactants. *Ther Deliv*, 4(5): 567-72, 2013.
- 65 How cheap will biosimilars need to be Posted. 0 5 / 0 8 / 2 0 1 1 , <http://gabionline.net/Biosimilars/Research/How-cheap-will-biosimilars-need-to-be>.