

Absorption enhancing excipients in systemic nasal drug delivery.

Maggio, ET*

Aegis Therapeutics LLC, San Diego, CA USA

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ABSTRACT

Intranasal drug delivery is becoming an increasingly important form of drug administration for chronic and chronic-intermittent diseases. Important new applications currently in development include drugs for diabetes, osteoporosis, obesity, certain types of convulsive disorders, migraine headaches, symptomatic pain relief, nausea, and anxiety, among others. Transmucosal absorption across the nasal mucosa is generally limited to molecules less than 1,000 Da. Systemic delivery of larger molecules requires formulations with a suitable transmucosal absorption enhancer. More than one hundred potential transmucosal absorption enhancing excipients have been tested to date. Nearly all have failed due to poor effectiveness or unacceptable toxicity to the mucosal tissue. Alkylsaccharides, cyclodextrins, and chitosans have emerged as leading candidates for potential broad clinical applications allowing the development of convenient, patient-friendly, needle free formulations of small molecule drugs, as well as, peptide and protein drugs that can be administered at home, at work, or in other public and private settings outside of the doctor's office or hospital environment.

KEY WORDS: Absorption enhancer, intranasal, drug delivery, chitosan, cyclodextrin, alkylsaccharide

INTRODUCTION

Intranasal drug delivery continues to evolve as an important form of drug delivery. The key aspects driving the growing adoption of nasal delivery include rapid drug absorption compared to subcutaneous injection or oral administration, the avoidance of hepatic first

pass effect, greater patient convenience and compliance, and the elimination of needle stick injuries and bio hazardous waste disposal problems associated with the use of syringes. In addition, consumer demographic trends point to an increasing popularity of self-administration of drugs for personal management and the control of chronic diseases. Topical nasal applications such as therapies for nasal congestion and allergic rhinitis still account for the major share of

* Corresponding author: Edward T. Maggio, PhD, Aegis Therapeutics LLC, San Diego, CA USA,
E-mail: emaggio@aegisthera.com

intranasal drug delivery. However, nasal administration of drugs intended for systemic absorption in the treatment of chronic diseases such as diabetes, osteoporosis, obesity, certain types of convulsive disorders, migraine headaches, together with symptomatic relief of pain, nausea, and anxiety is rapidly growing. The US market alone for intranasal drug delivery is projected to reach US \$4.4 billion by the year 2015.

ADVANTAGES OF INTRANASAL DELIVERY AND SOME EXAMPLES

Nasal delivery is suitable for the treatment of chronic conditions such as diabetes, osteoporosis, obesity, as well as, chronic-intermittent conditions such as migraine or breakthrough seizures. In general, intranasal delivery provides a convenient, patient-friendly, nonthreatening, needle free, pain free, administration route allowing for self-administration at home, at work, or in other public settings. As an example of improved convenience, at present the only approved out-of-hospital treatment for breakthrough seizures, in the US, is a diazepam rectal gel (Diastat®). Because of its inconvenient administration route it is prescribed essentially only for infants or very young children. Affected adults must wait for emergency treatment to arrive, transportation to hospital, intravenous administration of diazepam together with an overnight observational stay at a total cost of approximately US \$3,000, or more. A novel formulation, developed by San Diego-based Neurelis Corporation, incorporating a highly effective absorption enhancer has allowed the otherwise poorly nasally absorbed diazepam to be delivered with 96% absolute systemic bioavailability (1). As a replacement for rectal gel, it allows immediate treatment on-site and provides an excellent illustration of the importance of nasal delivery as a convenient alternate delivery mode.

An example of the benefit of increased speed of systemic absorption that can be achieved through nasal delivery is shown in Figure 1.

Figure 1 shows a comparison of the pharmacokinetics of nasal administration of sumatriptan containing the specific absorption enhancer dodecyl maltoside (DDM) with nasal administration of the same dose of sumatriptan in the absence of DDM. The T_{max} of nasally administered sumatriptan in the presence of the absorption enhancer is approximately 8 minutes compared to approximately 60-120 minutes for the currently available nasally administered sumatriptan. The DDM-enhanced nasal formulation reaches an equivalent therapeutic blood level in only 2-3 minutes, approximately 20-30 times faster than currently commercially available oral or nasal products (2).

Intranasal delivery can the speed absorption of much larger, otherwise injectable-only peptide drugs, as well. For peptides or proteins such as insulin, leptin, or growth hormone the T_{max} values for nasal administration have been shown to be one half or less of the corresponding T_{max} values for injections of these same proteins (3, 4).

Avoiding the hepatic first-pass metabolism effect is another advantage of systemic nasal delivery. Bypassing the GI tract allows more reliable and reproducible bioavailability to be achieved. Drugs that undergo extensive first-pass metabolism, display erratic absorption, or require quick therapeutic onset are potentially good drug candidates for intranasal delivery, particularly those that would otherwise require an injection.

NASAL ANATOMY, PHYSIOLOGY, AND ABSORPTION PROCESSES

The nose is divided into two nasal cavities by the septum, each with a volume of approximately 7.5 ml and a surface area of approximately 75 cm² (5, 6). Of the three distinct functional regions in the nose, namely, the vestibular, respiratory, and olfactory regions, the respiratory region is the largest and comprises approximately 65 cm². It is highly vascularized and is the principal site of systemic drug absorption (6). The respiratory epithelium

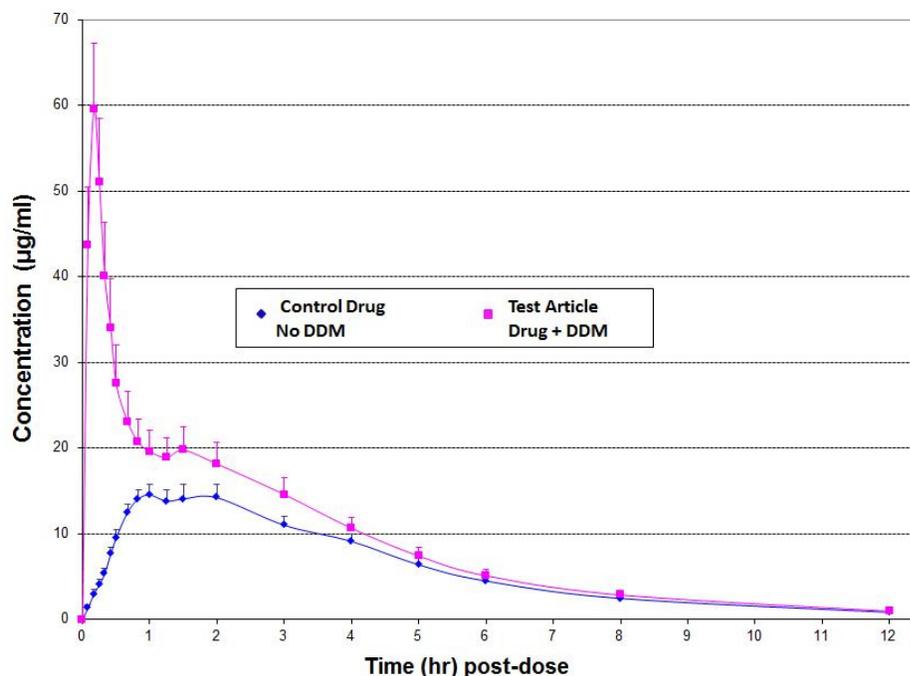


Figure 1 The absorption enhancement effect of DDM on the plasma profile of nasally administered sumatriptan in humans (mean \pm S.D.). T_{max} is reduced from 60-120 for the currently marketed nasal or oral sumatriptan down to 8 minutes with equivalent therapeutic levels achieved at 2 minutes, and the bioavailability as measured by the AUC (area under the curve) is significantly increased.

consists of basal cells, mucus-containing goblet cells, ciliated columnar cells, and non-ciliated columnar cells (5, 6). The cilia are surrounded by a film of mucus and move in a continuous wavelike fashion to transport mucus and entrapped particles to the pharynx for ingestion (6 - 8).

Mucus is a viscous colloid comprised of mucins (glycoproteins that are produced by goblet cells in the mucous membranes and submucosal glands), together with antibacterial proteins such as lysozyme and lactoferrin, immunoglobulins, and inorganic salts (9). The constantly regenerating mucus coating moves at 5 to 6 mm/min. (5, 10) and serves to protect epithelial cells from external insults (viruses, bacteria, and chemical irritants). By restricting drop size in nasally administered sprays to a diameter $>10 \mu\text{m}$ deposition is restricted to the nasal cavity and lung exposure is essentially zero (11-14). The pH range of the nasal cavity

is approximately 5.5 to 6.5 so nasal irritation is minimized when products are formulated within this pH range (15, 16). The total spray volume that can be reliably delivered to each naris is limited by the size of the nasal cavity and generally thought to be no more than 150 μl and the upper limit of a drug dose has been suggested to be 25 mg/dose (16).

ABSORPTION ENHANCING EXCIPIENTS

There are two primary mechanisms for absorption through the mucosa. They are paracellular transport via opening of tight junctions between cells, and transcellular transport or transcytosis through cells via vesicle carriers (3, 17). Obstacles to drug absorption are potential metabolism before reaching the systemic circulation and, limited residence time in the cavity. Some transmucosal absorption enhancers function by altering either or both the paracellular and transcellular

pathways, while others serve to extend residence time in the nasal cavity or prevent metabolic changes, e.g., unwanted peptide hydrolysis.

Dozens of excipients have been tested as potential enhancers of intranasal absorption over the course of the last three decades in the hope that alternate noninvasive means of administering peptides, proteins, and small molecule drugs might be achieved. Table 1 lists some of these. Nearly all of the molecules tested to date have been shown to cause significant and, in some cases, serious damage to the nasal mucosa upon repeated administration, especially at concentrations high enough to achieve a substantial degree of transmucosal absorption enhancement. Indeed, the principal factors limiting broad acceptance of intranasal administration have been damage to the nasal mucosa coupled with poor transmucosal absorption caused by the absorption enhancers. The few exceptions, which include the alkylsaccharides, cyclodextrins, and chitosans, are among the most effective and are generally well tolerated, with varying degrees of effectiveness as absorption enhancers.

In the absence of an absorption enhancer, the apparent molecular weight cut-off point for nasal absorption is approximately 1,000 Da, with molecules less than 1,000 Da having better absorption (43). Co-administration of drugs with absorption enhancers promotes the absorption of drugs ranging in size from 1,000 Da to 31,000 Da. The principle mechanisms for increased absorption are loosening of tight junctions between cells, enhanced vesicular transcytotic transport, alteration of the rheological (fluidity) properties of the mucus and/or alteration of the cilia (i.e., paralysis of ciliary beating or removal of cilia from the epithelial cells that line the nasal cavity (3, 23, 44-52). The latter two mechanisms serve to increase the residence time of drug in the nasal cavity allowing more time for passive diffusion.

Table 1 Some examples of molecules that have been used as transmucosal absorption enhancers. Excipients used in human trials are shown in **bold**. Of these, excipients exhibiting no reported irritation are marked with an asterisk (*).

Aprotinin (18)
Benzalkonium chloride* (19)
Benzyl Alcohol (20)
Capric acid, sodium salt (21)
Ceramides (22)
Cetylpyridinium chloride (19)
Chitosan* (23)
Cyclodextrins* (22)
Deoxycholic acid (24)
Decanoyl carnitine (21)
Dodecyl maltoside* (4)
EDTA (21)
Glycocholic acid , sodium salt (25)
Glycodeoxycholic acid, sodium salt (25)
Glycofurol (26)
Glycosylated sphingosines (22)
Glycyrrhetic acids (27)
2-Hydroxypropyl- β -cyclodextrin (28)
Laureth-9 (29)
Lauric acid (30)
Lauroyl carnitine (31)
Lauryl sulfate, sodium salt (25)
Lysophosphatidylcholine (29)
Menthol (32)
Poloxamer 407 (33)
Poloxamer F68 (34)
Poly-L-arginine (35)
Polyoxyethylene-9-lauryl ether (25)
Polysorbate 80 (34)
Propylene glycol (36)
Quillaja saponin (37)
Salicylic acid, sodium salt (20)
β -Sitosterol- β -D-glucoside (38)
Sucrose cocoate (39)
Taurocholic acid , sodium salt (40)
Taurodeoxycholic acid, sodium salt (41)
Taurodihydrofusidic acid , sodium salt (42)
Tetradecyl maltoside (2)

TOXICITY ASSESSMENT

Two principal requirements for useful absorption enhancers are significant effectiveness and safety. Extensive *in vivo* studies aimed at assessing the effectiveness and

toxicity of more than 50 possible absorption enhancers were conducted by Chen *et al.* (22) using the measurement of transepithelial electrical resistance across confluent cell layers comprised of cultured human bronchial/tracheal epithelial cells in a microtiter well format. If exposure to the cells by one of the 50, or so, test excipients resulted in a reduction in transepithelial electrical resistance (TEER) across the confluent cell layer, it was interpreted as an opening of tight junctions between the cells. Similar studies have been repeated by Vllasaliu *et al.* using Calu-3 cells (53). The reduction in TEER *in vitro* seems to correlate with potential absorption enhancement properties *in vivo* though the correlation is not by any means perfect. Attempts to use this same cell based model for toxicity assessment is not predictive because of gross difference between the *in vitro* and *in situ* cell environments. *In vitro* cell culture models of nasal mucosa are understandably deficient from a number of perspectives. Some of these deficiencies apply in general to assessment of all excipients and some are deficiencies specific to the particular excipient being tested. From a general deficiency perspective, both studies exposed the naked confluent cells in culture media to excipients for 60 to 120 minutes at 37°C. During this time, the naked cells were continuously exposed to continuous high concentrations of the excipient.

In contrast, in the nasal cavity, the epithelial cells lining the nasal cavity are bathed in a constantly regenerating mucus coating that flows at a rate of 5 - 6 mm/min. resulting in a mucociliary clearance time of approximately 15 minutes. Thus the contact time of exposure of nasal epithelia, and the concentration of both excipient and drug is progressively and continuously reduced by three means i.e., dilution into the mucus, absorption into the systemic circulation, and physical removal through the mucociliary clearance mechanism. Assuming a linear concentration reduction over 15 minutes the integrated exposure (i.e., time X concentration) to excipient is lower by a factor of 8- to 16-fold in the physiological

circumstance compared to the 60 to 120-minute incubation times, respectively, for the naked cells. Further, the intrinsic properties of mucus whose complex composition of mucin glycoproteins, lysozyme, lactoferrin, and immunoglobulins, is precisely designed as a natural protection against external insults cannot be factored into the *in vitro* cell assay.

ALKYLSACCHARIDES

Alkylglycosides and sucrose esters of fatty acids are nonionic alkylsaccharide surfactants that consist of an aliphatic hydrocarbon chain coupled to a sugar moiety by a glycosidic or ester bond, respectively. The particular alkylsaccharides that were shown to be effective absorption enhancers are odorless, tasteless, non-toxic, non-irritating, non-mutagenic, and non-sensitizing in the Draize test at concentrations of up to 25% (54, 55). They are synthesized as single chemical entities composed of a sugar, typically a disaccharide, and an alkyl chain, typically 10 to 16 carbon atoms in length. They provide controlled transient permeation of the nasal mucosal barrier with no irritation. Following absorption or ingestion they metabolize to CO₂ and H₂O through the corresponding sugar and fatty acid (56, 57).

Alkylglycosides are widely used in the food industry. For example, they are sprayed on fruits and vegetables to prevent the growth of bacteria and fungi or, used to clean food processing equipment, primarily because of their intrinsic and highly effective antimicrobial activity and lack of toxicity. The EPA has determined that there is no need to establish an upper limit of exposure for adults, children or infants (58).

Similarly, sucrose esters find widespread use as food-grade emulsifiers and in cosmetic preparations. The No Observed Effect Level (NOEL) for these molecules is as high as 2,000 mg/kg body weight in some instances (56) and are designated as Generally Recognized As Safe (GRAS) substances for food applications. They

are used in such small amounts that the WHO oral allowable daily intake (ADI) is nearly 10,000 times (56) the amount that would be used in a typical single nasal spray dose (i.e., 200 µg/spray *versus* 2 g/day ADI).

Despite the accumulation of considerable experimental evidence concerning the efficacy of alkylglycosides as absorption-enhancers, their mechanism of action remains unknown. Alkyl maltosides appear to enhance transmucosal delivery of peptides through transcellular, and paracellular, pathways (3, 59). Transmission electron micrographs of the nasal septa of rats show, immediately after the exposure to alkyl maltoside, unstained regions that are consistent with cellular internalizations and areas of thinned cilia associated with vesicle formation (3). The transcellular pathway is further supported by fluorescence light micrographs showing the internalization of fluorescein-labeled insulin administered intranasally with an alkyl maltoside (3). As stated above, the paracellular pathway is nominally demonstrated by a decrease in the transepithelial electrical resistance (TEER) and an increase in mannitol movement across a confluent layer of human bronchial epithelial cells in the presence of tetradecyl maltoside (36).

Studies have shown that alkylglycosides and sucrose esters are among the most effective nasal absorption enhancers for a wide range of peptide, protein, and non-peptide macromolecular drugs in rats, mice, cats, dogs and monkeys (60-66). Dose-escalation studies were conducted in rats to determine the potencies of various alkylglycosides and sucrose esters in increasing nasal absorption of insulin and to determine the contribution of the alkylchain and the sugar moiety. These studies revealed that shorter chained alkylsaccharides coupled to glucose, such as hexyl, heptyl, octyl or nonyl glucoses, were ineffective, or minimally effective, at promoting insulin absorption from the nose (63, 65).

Intermediate-length alkylsaccharides such as decanoyl sucrose, decyl maltoside, or octyl maltoside were more effective in promoting nasal insulin absorption. Longer chain alkylsaccharides such as dodecyl maltoside, tridecyl maltoside, tetradecyl maltoside, and sucrose dodecanoate were very potent absorption enhancers, even at concentrations as low as 0.03-0.06%. No other absorption-enhancing agents tested to date have been as effective at such low concentrations. Interestingly, increasing the alkyl chain length beyond 14 carbons (e.g., pentadecyl maltoside, or hexadecyl maltoside) decreases the potency of nasal insulin absorption.

CYCLODEXTRINS

Cyclodextrins (CD) are cyclic oligosaccharides composed of six or more monosaccharide units with a central cavity. Cyclodextrins can form inclusion complexes with hydrophobic molecules and they have primarily been used to increase drug solubility and dissolution and to enhance low molecular weight drug absorption (67, 68). Among the cyclodextrin derivatives studied as potential nasal insulin absorption enhancers, dimethyl-beta-cyclodextrin was found to be the most effective, while alpha-CD was less effective and beta- and gamma-CD had negligible effects on insulin absorption (69). Cyclodextrins are believed to interact with cholesterol within the cell membrane (70). This interaction transiently opens tight junctions which may explain their ability to facilitate peptide absorption across the nasal mucosa (49). Since not all cyclodextrins are effective at increasing peptide bioavailability, the architecture of the central cavity appears to be critical for nasal peptide drug absorption. Previous work has demonstrated that dimethyl-beta-cyclodextrin was effective in promoting the absorption of a number of drugs including insulin (72, 73), calcitonin (40), and low molecular weight heparin (74). Gamma cyclodextrin is ineffective at increasing peptide drug absorption.

CHITOSAN

Chitosan is a linear cationic polysaccharide produced from the deacetylation of chitin, a component of the shells of shrimp and other crustaceans (7, 79, 80). Chitin is composed of randomly distributed β -(1-4)-linked D-glucosamine and N-acetyl-D-glucosamine joined by glycosidic bonds.

Chitosan has been shown to increase the bioavailability of insulin and other small peptides and polar macromolecules in different animal models (75-77). Chitosan exhibits bioadhesive properties and interacts strongly with nasal mucus layer enhancing the contact time for drug with membrane. The addition of 0.2-0.5% chitosan to nasal formulations of insulin resulted in significant increases in plasma insulin and reductions in blood glucose in both sheep and rat models. Studies in human volunteers demonstrated that nasal administration of chitosan formulations results in significantly longer nasal clearance times (78) suggesting that chitosan decreases mucociliary clearance and prolongs the residence time of peptides within the nasal cavity (79-86). Chitosan also produces a transient opening of tight junctions of confluent Caco-2 cells (87).

BILE SALTS AND DERIVATIVES

Attempts to administer insulin non-invasively through the nasal route using bile salts date back nearly three decades (42) with mixed results. Bile salts and their derivatives, such as sodium glycocholate, sodium taurocholate, and sodium taurodihydrofusidate, were shown to effectively promote nasal insulin absorption (41, 45, 88, 89) and were subsequently extensively studied for their ability to promote the absorption of a variety of peptide drugs from several alternative delivery sites (90, 91). The mechanisms of action by which the bile salts/derivatives promote increased nasal absorption of peptide drugs are not well understood. Possible explanations may include the fluidization of the nasal epithelial cell membranes, increases in transcytotic movement

of peptides via endocytotic vesicles, or the inhibition of certain proteolytic enzymes capable of degrading peptides before they can successfully cross the nasal epithelium. Inclusion of sodium glycholate in an insulin formulation resulted in a significant reduction in enzymatic degradation of insulin (92). Through circular dichroism and alpha-chymotrypic degradation studies, a dose-response relationship between increasing concentrations of sodium glycholate and the presence of monomeric insulin has been shown (92). Similarly, sodium taurocholate has been shown to increase disaggregation of insulin hexamers in a dose dependent manner (92). Unfortunately, in test subjects bile salts and derivatives were found to produce significant nasal irritation, stinging and lacrimation. So while early attempts to administer insulin using bile salt-based formulations successfully achieved useful systemic blood levels and a corresponding reduction in blood glucose levels, the resulting nasal damage was unacceptable and not tolerated by test subjects. This experience with the bile salt excipients illustrates the most significant challenge for the practical broad adoption of nasal drug delivery.

COMPARATIVE STUDIES

The three types of absorption enhancers that have been studied most extensively, namely alkylsaccharides, cyclodextrins, and chitosan, have proven useful for nasal delivery of small molecule drugs, as well as, for peptide and protein drugs. Each offers potential advantages over the others in specific applications. For example, chitosan has proven useful as an absorption enhancer for both aqueous solution and dry powder formats. Cyclodextrins have been used in aqueous solutions where they not only increase transmucosal absorption, but can also increase solubility of hydrophobic drugs. Alkylsaccharides, which provide the greatest degree of absorption enhancement in most applications, are soluble in aqueous and oil-based/organic liquid formulations and are being used in both formats. In addition, alkylsaccharides prevent peptide and protein

Table 2 Comparison of the absolute bioavailability for intranasally administered calcitonin using three different absorption enhancers.

TEST ARTICLE	EXCIPIENT	ABSOLUTE BIOAVAILABILITY	TOTAL DOSE ADMINISTERED	REFERENCE
Intravenous calcitonin control	none	100%	10 IU/KG	
Nasal calcitonin, in pH 4 isotonic phosphate buffer	1% chitosan free amine	2.45%	10 IU/KG	
Nasal calcitonin, in pH 4 isotonic phosphate buffer	5% dimethyl-beta-cyclodextrin	1.91%	10 IU/KG	(28)
Nasal calcitonin in pH 4 isotonic phosphate buffer control	None	1.22%	10 IU/KG	
Nasal calcitonin in pH 3.75, 6 mM sodium acetate, 0.9% sodium chloride	0.125% tetradecyl maltoside	52%	8 IU/KG	(66)

aggregation (93, 94). There are few direct comparative studies assessing differences in absorption enhancement efficiency in the literature. Published studies on the nasal bioavailability of calcitonin, a peptide drug 32 amino acids in length with an approximate molecular weight of 3,500 Da (66, 93) provides one such example. Sinswat *et al.* (28) compared the absolute bioavailability of calcitonin administered nasally to rats using cyclodextrins and chitosan's as the absorption enhancers to intravenous calcitonin as the control. Ahsan *et al.* carried out a similar study, again in rats, using alkylsaccharide tetradecylmaltoside as the absorption enhancer, again using intravenous administration as the control (66). The results are shown above in Table 2.

In these side by side comparisons, the formulation containing the alkylsaccharide excipient tetradecyl maltoside was found to be significantly more effective than either of the formulations containing chitosan or dimethyl beta cyclodextrin.

Data from a study comparing intranasal absorption of a GLP-1 analog oligopeptide in Sprague Dawley rats are shown in Figure 2. In this study, two alkylsaccharides, dodecyl maltoside (DDM) and sucrose dodecanoate (SDD) were compared to the bile acid absorption enhancer sodium taurocholate, together with Tween 20 (polysorbate-20) and phosphate buffered saline as a negative control. Rats were anesthetized using isoflurane/O₂ mixture and dosed with 40 µg of the GLP-1

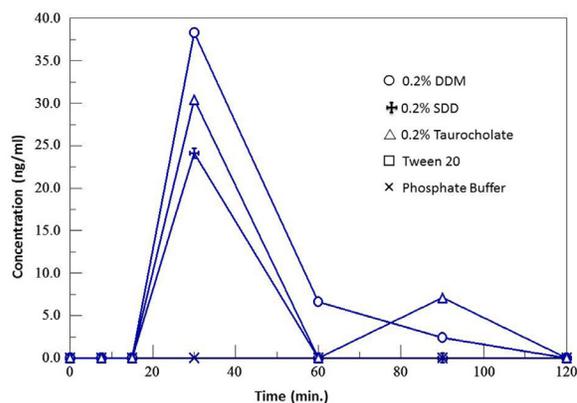


Figure 2 Single dose pharmacokinetics study of intranasally administered GLP-1 analog in Sprague Dawley rats. Data courtesy of QED Bioscience Inc., San Diego, CA.

analog by instillation of a 20 µl drop into a single naris. Solutions of the GLP-1 analog and excipient (DDM, SDD, sodium taurocholate, Tween-20) were prepared in 20 mM phosphate buffer, pH 7.0. Each group of 12 rats was matched by body weight. Blood samples were drawn at the timed intervals shown and assayed.

The molecular weight of GLP-1 at 4,112 Da is slightly larger than calcitonin, but very much within the molecular weight range suitable for intranasal peptide administration. Of the two alkylsaccharides tested, dodecyl maltoside appeared to be better than sucrose dodecanoate and both were superior to sodium taurocholate. The polysorbate surfactant Tween-20 exhibited no absorption enhancement and was comparable to the phosphate buffer control.

CONCLUSION

More than one hundred potential transmucosal absorption enhancing excipients have been tested to date. Nearly all failed due to poor effectiveness or unacceptable toxicity to mucosal tissue. Alkylsaccharides, cyclodextrins, and chitosan's have emerged as the leading candidates for potential broad clinical applications. Certain molecules from all three types are able to exert their desired effects without causing transmucosal damage and all three have demonstrated clinical utility in human studies together with demonstrated safety and lack of intranasal toxicity. A principal mechanistic benefit shared by these absorption enhancing excipients is the fact that they function independently of the drug, that is, they act on the transmucosal membranes and do not require a modification of the drug substance.

It is certainly desirable that additional research is carried out to identify better alternatives. In undertaking such research there are two impediments that must be overcome. First, there is a practical limit to the size of molecules that can be absorbed transmucosally. A practical molecular weight limit is estimated to be in the range of approximately 25 – 30 kDa. This is because there is a physical limit to the extent to which tight junctions can be opened without tissue damage. Absorption by the transcellular route is not subject to similar size limitation and may even allow absorption of nanoparticles, well beyond the size of soluble peptide or protein molecules (95). It is possible that a combination of excipients which separately maximize paracellular and transcellular absorption could be worth pursuing. Nevertheless, it may be difficult to develop alternative excipients that exceed the performance of those tested to date without causing unacceptable tissue damage.

A second impediment and practical “real world” concern relates to the regulatory requirement to assess the potential for toxicity before being permitted to conduct human clinical trials. In every case, care must be taken to fully assess the potential for toxicity upon chronic administration of novel excipients.

Today, literally thousands of patients have been exposed to intranasal administration of drugs using alkylsaccharides, cyclodextrins, and chitosan's and thus there is a growing database of safety experience with these excipients. Because of the ever-increasing cost of preclinical, as well as, clinical studies the cost to duplicate this experience base in itself has become somewhat of a significant impediment.

REFERENCES

1. Neurelis announces positive results from phase 1 pharmacokinetic study of NRL-01 (intranasal diazepam) <http://www.prnewswire.com/news-releases/neurelis-announces-positive-results-from-phase-1-pharmacokinetic-study-of-nrl-01-intranasal-diazepam-123834469.html> (accessed 17 April 2014).
2. Maggio ET. Alkylglycoside compositions for drug administration. US Patent No. 8,268,791, Issued Sept. 18, 2012.
3. Arnold JJ, Ahsan F, Meezan E, Pillion DJ. Correlation of tetradecylmaltoside induced increases in nasal peptide drug delivery with morphological changes in nasal epithelial cells. *J. Pharm. Sci.*, 93(9):2205–13, 2004.
4. Maggio ET, Meezan E, Ghambeer DKS, Pillion DJ. High bioavailability formulation of salmon calcitonin—potential opportunities for expanded use in analgesia. *Drug Deliv. Technol.*, 10:58–63, 2010.
5. Mygind N, Ånggård A., Anatomy and physiology of the nose – pathophysiologic alterations in allergic rhinitis. *Clin Rev Allergy*. 2:173-188, 1984.
6. Illum L., Transport of drugs from the nasal cavity to the central nervous system. *Eur J Pharm Sci.*, 11(1):1-18, 2000.
7. Schipper NGM., Verhoef JC, Merkus FWHM. The nasal mucociliary clearance: relevance to nasal drug delivery. *Pharm Res.*, 8:807-814, 1991.
8. Mathison S, Nagilla R, Kompella UB., Nasal route for direct delivery of solutes to the central nervous system: fact or fiction? *J Drug Target*. 5:415-441, 1998.
9. Singh PK, Parsek MR, Greenberg EP, Welsh MJ. A component of innate immunity prevents bacterial biofilm development. *Nature*, 417 (6888): 552–5, 2002.
10. Chien YW, Su KSE, Chang S. *Nasal Systemic Drug Delivery*. New York: Marcel Dekker, Inc, 1989:1-38.
11. Rothe H, Fautz R, Gerber E, Neumann L, Rettinger K, Schuh W, Gronewold C. Special aspects of cosmetic spray safety evaluations: Principles on

- inhalation risk assessment. *Toxicol. Lett.*, 205(2):97-104, 2011.
12. Hatch TF. Distribution and deposition of inhaled particles in respiratory tract. *Bacteriol.Rev.* 25:237-240, 1961.
 13. Heyder J, Gebhart J, Rudolf G, Schiller CF, Stahlhofen W. Deposition of particles in the human respiratory tract in the size range 0.005-15 μm . *J.Aerosol.Sci.*, 17(5):811-825,1986.
 14. Oberdorster G, Oberdorster E, Oberdorster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ.Health Perspect.*, 113(7):823-839, 2005.
 15. England RJ, Homer JJ, Knight LC, Ell SR. Nasal pH measurement: a reliable and repeatable parameter. *Clin. Otolaryngol. Allied Sci.*, 24(1):67-8, 1999.
 16. Behl CR, Pimplaskar HK, Sileno AP, deMeireles J, Romeo VD. Effects of physicochemical properties and other factors on systemic nasal drug delivery. *Adv. Drug Delivery Rev.*, 29:89-116, 1998.
 17. Martin E, Verhoef JC, Cullander C, Romeijn SG, Nagelkerke JF, Merkus FWHM. Confocal laser scanning microscopic visualization of the transport of dextrans after nasal administration to rats: effects of absorption enhancers. *Pharm Res.*, 14:631-637, 1997.
 18. Morimoto K, Yamaguchi H, Iwakura Y, Miyazaki M, Nakatani E, Iwamoto T, Ohashi Y, Nakai Y. Effects of proteolytic enzyme inhibitors on the nasal absorption of vasopressin and an analogue. *Pharm. Res.* 18(9):1175-9, 1991.
 19. Green K. 1993 The effects of preservatives on corneal permeability of drugs. In *Biopharmaceutics of Ocular Drug Delivery*, ed Edman P. 43 Boca Raton, FL: CRC Press..
 20. Kagatani S, Hasumi S, Sonobe T, Aruga M. Pharmaceutical compositions for nasal administration comprising calcitonin and an absorption-promoting substance US Patent No. 4,690,952 (1987).
 21. Tomita, M., Hayashi M, Awazu.S. Absorption-enhancing mechanism of EDTA, caprate, and decanoylcarnitine in Caco-2 cells. *J. Pharm. Sci.* 85(6): 608-611, 1996.
 22. Chen-Quay S-C, Eiting KT, Li A, Lamharzin, Quay SC. Identification of Tight Junction Modulating Lipids. *J Pharm Sci.*, 98(2):606-19, 2009.
 23. Davis SS, Illum I. Absorption enhancers for nasal drug delivery. *Clin. Pharmacokinet.* 42(13): 1107-1128, 2003.
 24. Chavanpatil MD, Vavia PR. Nasal drug delivery of sumatriptan succinate. *Pharmazie.* 60(5):347-9, 2005.
 25. Donovan, MD, Flynn GL, Amidon GL. The Molecular Weight Dependence of Nasal Absorption: The Effect of Absorption Enhancers." *Pharmaceutical Research* 7 (8): 808-815, 1990.
 26. Bagger MA, Nielsen HW, Bechgaard E. Nasal bioavailability of peptide T in rabbits: absorption enhancement by sodium glycocholate and glycofuroil. *Eur. J. Pharm. Sci.* 14(1):69-74, 2001.
 27. Mishima M, Okada S, Wakita Y, Nakano M. Promotion of nasal absorption of insulin by glycyrrhetic acid derivatives. I. *J. Pharmacobiodyn.* 12(1):31-6, 1989.
 28. Sinswat P, Tengamnuy P. Enhancing effect of chitosan on nasal absorption of salmon calcitonin in rats: comparison of hydroxyl propyl and dimethyl-beta-cyclodextrins. *Int. J. Phar.* 257:15-22, 2003.
 29. Aungst BJ, Rogers NJ. Site dependence of absorption-promoting actions of laurith-9, Na salicylate, Na2EDTA, and aprotinin on rectal, nasal, and buccal insulin delivery. *Pharm. Res.* 5:305-8, 1998.
 30. Lindmark T, Kimura Y, Artursson P. Absorption enhancement through intracellular regulation of tight junction permeability by medium chain fatty acids in Caco-2 cells. *J. Pharmacol. Exp. Ther.* 284(1):362-9, 1998.
 31. Kagatani S, Shinoda T, Fukui M, Ohmura T, Hasumi S, Sonobe T. Enhancement of nasal salmon calcitonin absorption by lauroylcarnitine chloride in rats. *Pharm. Res.* 13(5):739-43, 1996.
 32. Wang H, Xu W, M.Effect of l-menthol pretreated nasal cavity on insulin pharmacological bioavailability. *Chinese Pharmacological Bulletin*;2002-01, 2002.
 33. Parmar VJ, Lumbhani AN. Formulation and development of thermo- reversible mucoadhesive intranasal in situ hydrogel by using a combination of polymers. *Bull. Pharmaceut. Res.* 2(3):167-74, 2012.
 34. Lin H, Gebhardt M, Bianc S, Kwond KA. , Shima C-K, Chunga S-J, Kima D-D. Enhancing effect of surfactants on fexofenadine•HCl transport across the human nasal epithelial cell monolayer. *Int. J. Pharmaceut.* 330: 23–31, 2007.
 35. Ohtake K, Natsume H, Ueda H, Morimoto Y. Analysis of transient and reversible effects of poly-L-arginine on the in vivo nasal absorption of FITC-dextran in rats. *J. Control. Release.* 82(2-3):263-75, 2002.
 36. Williams AC, Barry BW. Penetration enhancers. *Adv. Drug Deliv. Rev.* 56(5):603-18, 2004.
 37. Pillion DJ, Amsden JA, Kensil CR, Recchia J. Structure-function relationship among Quillaja

- saponins serving as excipients for nasal and ocular delivery of insulin. *J. Pharm. Sci.* 85(5):518-24, 1996.
38. Ando T, Maitani Y, Yamamoto T, Takayama K, Nagai T. Nasal insulin delivery in rabbits using soybean-derived sterylglucoside and sterol mixtures as novel enhancers in suspension dosage forms. *Biol. Pharm. Bull.* 21(8):862-5, 1998.
 39. Ahsan F, Arnold JJ, Meezan E, Pillion DJ. Sucrose cocoate, a component of cosmetic preparation, enhances nasal and ocular peptide absorption, *Int. J. Pharm.*, 251:195-203, 2003.
 40. Yetkin G, Celebi N, Agabeyoglu I, Gokcora N. The effect of dimethyl-beta-cyclodextrin and sodium taurocholate on the nasal bioavailability of salmon calcitonin in rabbits, *STP. Pharma. Sci.*, 9:249-53, 1999.
 41. Gordon GS, Moses AC, Silver RD, Flier JS, Carey MC. Nasal absorption of insulin: enhancement by hydrophobic bile salts. *Proc. Natl. Acad. Sci. USA*, 82(21): 7419-7423, 1985.
 42. Baldwin PA, Klingbeil CK, Grimm CJ, Longenecker JP. The effect of sodium tauro-24,25-dihydrofusidate on the nasal absorption of human growth hormone in three animal models. *Pharm. Res.*, 7(5):547-552, 1990.
 43. McMartin C, Hutchinson LEF, Hyde R, Peters GE. Analysis of structural requirements for the absorption of drugs and macromolecules from the nasal cavity. *J Pharm Sci.*, 76:535-540, 1987.
 44. Illum L. Nasal drug delivery: new developments and strategies. *Drug Discov. Today*, 7:1184-89, 2002.
 45. Aungst BJ, Rogers NJ, Shefter E. Comparison of nasal rectal buccal, sublingual, and intramuscular insulin in efficacy and the effects of a bile salt absorption enhancer. *J. Pharm. Exp. Ther.*, 244:23-27, 1988.
 46. Zhang Y, Zhang Q, Sun Y, Sun J, Wang X, Chen M. Nasal recombinant hirudin-2 delivery: absorption and its mechanism in vivo and in vitro studies. *Biol. Pharm. Bull.* 28(12): 2263-2267, 2005.
 47. Donovan, MD, Huang Y. Large molecule particulate uptake in the nasal cavity: the effect of size on nasal absorption. *Adv. Drug Del. Rev.*, 29, 147-155, 1998.
 48. Lee VH, Yamamoto A, Kompella UB. Mucosal penetration enhancers for facilitation of peptide and protein drug absorption. *Crit. Rev. Ther. Drug Carrier Syst.*, 8, 91- 92, 1991.
 49. Wermeling DP, Miller JL, Rudy AC. Systemic intranasal drug delivery: concepts and applications. *Drug Deliv. Tech.*, 2, 56, 2002.
 50. Khanvilkar K, Donovan MD, Flanagan DR. Drug transfer through mucus. *Adv. Drug Deliv. Rev.*, 48:173-93, 2001.
 51. Lutz KL, Siahaan TJ. Molecular structure of the apical junction complex and its contribution to the paracellular barrier. *J. Pharm. Sci.*, 86, 977-84, 1997.
 52. Agerholm C, Bastholm L, Johansen PB, Nielsen MH, Elling F. Epithelial transport and bioavailability of intranasally administered human growth hormone formulated with the absorption enhancers didecanoyl-L-alpha-phosphatidylcholine and alpha-cyclodextrins in rabbits. *J. Pharm. Sci.*, 83, 1706-11, 1994.
 53. Vllasaliu D, Shubber, S, Fowler R, Garnett M, Alexander C, Stolnik S, Epithelial toxicity of alkylglycosides surfactants. *J. Pharm. Sci.*, 102(1):114-125, 2013.
 54. Material Safety Data Sheet TEGOSOFT LSE 65 K SOFT Evonik Goldschmidt GmbH Version: 1.11 Date Issued: 01/21/2010 Gold-schmidtstr. 100 Essen, 05 45127.
 55. Kocher K, Wiegand HJ. Toxicology and dermatology. In: Balzer D, editor. *Surfactant science series*, Vol. 91, Non ionic surfactants: alkylpolyglucosides. New York: Marcel Dekker; 2000. p. 365-83.
 56. Sucrose esters of fatty acids and sucroglycerides (WHO Food Additives Series 40), The forty-ninth meeting of the Joint FAO/ WHO Expert Committee on Food Additives (JECFA) Geneva 1998. <http://www.inchem.org/documents/jecfa/jecmono/v040je04.htm>.
 57. Weber N, Benning H. Metabolism of orally administered alkyl beta-glycosides in the mouse. *J. Nutr.*, 114(2):247-54, 1984.
 58. Federal register ref. CFR Part 180. Alkyl (C10-C16) polyglycosides; exemption from the requirement of a tolerance. Federal register 70(177), 54281-54286, 2005.
 59. Ahsan F, Arnold JJ, Yang T, Meezan E, Schwiebert EM, Pillion DJ. Effects of the permeability enhancers, tetradecylmaltoside and dimethyl-beta-cyclodextrin, on insulin movement across human bronchial epithelial cells (16HBE14o⁻). *European Journal of Pharmaceutical Sciences.*, 20:27-34, 2003.
 60. Pillion DJ, Hosmer S, Meezan E. Dodecylmaltoside-mediated nasal and ocular absorption of lyspro-Insulin: independence of surfactant action from peptide multimer dissociation. *Pharm. Res.*, 15:1637-39, 1998.
 61. Waldrop MA, Leinung MC, Lee DW, Grasso P. Intranasal delivery of mouse [D-Leu-4]-OB3, a synthetic peptide amide with leptin-like activity, improves energy balance, glycemic control, insulin

- sensitivity, and bone formation in leptin resistant C57BLK/6-m db/db mice. *Diabetes Obes. Metab.*, 12(10):871-5, 2010.
62. Ahsan F, Arnold JJ, Meezan E, Pillion DJ. Mutual inhibition of the insulin absorption-enhancing properties of dodecylmaltoside and dimethyl-beta-cyclodextrin following nasal administration. *Pharm. Res.*, 18: 608-14, 2001.
 63. Pillion, DJ, Atchison JA, Gargiulo C, Wang RX, Wang P, Meezan E. Insulin delivery in nosedrops: new formulations containing alkylglycosides. *Endocrinology*, 135:2386-91, 1994.
 64. Pillion DJ, Atchison JA, Meezan E. Alkylglycosides Enhance Systemic Absorption of insulin delivered topically to the rat eye. *J. Pharmacol. Exp. Therp.*, 271:1274-80, 1994.
 65. Pillion DJ, Ahsan F, Arnold JJ, Balusubramaniam BM, Pirana O, Meezan E. Synthetic long-chain alkylglycosides as enhancers of nasal insulin absorption. *J. Pharm. Sci.*, 91: 1456-62, 2002.
 66. Ahsan, F, Arnold J, Meezan E, Pillion DJ, Enhanced bioavailability of calcitonin formulated with alkylglycosides following nasal and ocular administration in rats. *Pharm. Res.*, 18: 1742-46, 2001.
 67. Rajewski RA, Stella VJ. Pharmaceutical applications of cyclodextrins 2. In vivo drug delivery. *J. Pharmaceut. Sci.*, 85:1142-69, 1996.
 68. Merkus FW, Verhoef JC, Martin E, Romeijn SG, van der Kuy PH, Hermens WA, Schipper NG. Cyclodextrins in nasal drug delivery, *Adv. Drug Del. Rev.*, 36:41-57, 1999.
 69. Merkus FW, Verhoef J C, Romeijn SG, Schipper NG. Absorption enhancing effect of cyclodextrins on intranasally administered insulin in rats. *Pharm. Res.*, 8:588-92, 1991.
 70. Francis SA, Kelly JM, McCormack J, Rogers RA, Lai J, Schneeberger EE, Lynch RD. Rapid reduction of MDCK cell cholesterol by methyl-beta-cyclodextrin alters steady state transepithelial electrical resistance. *Eur. J. Cell. Biol.*, 78:473-84, 1999.
 71. Martin E, Verhoef J C, Merkus FWHM. Efficacy, safety and mechanism of cyclodextrins as absorption enhancers in nasal delivery of peptide and protein drugs. *J. Drug Target*, 6:17-36, 1998.
 72. Shao Z, Krishnamoorthy R, Mitra K Cyclodextrins as nasal absorption promoters of insulin: mechanistic evaluations. *Pharm. Res.*, 9(9):1157-63, 1992.
 73. Watanabe Y, Matsumoto Y, Kawamoto K, Yazawa S, Matsumoto M. Enhancing effect of cyclodextrins on nasal absorption of insulin and its duration in rabbits. *Chem. Pharm. Bull. (Tokyo)*, 40(11):3100-3104, 1992.
 74. Yang T, Hussain A, Paulson J, Abbruscato TJ, Ahsan F. Cyclodextrins in nasal delivery of low-molecular-weight heparins: in vivo and in vitro studies. *Pharm. Res.*, 21:1127-36, 2004.
 75. Illum L, Dodane V, Iqbal K. Chitosan technology to enhance the effectiveness of nasal drug delivery. *Drug. Deliv. Tech.*, 2: 40, 2002.
 76. Illum L. Chitosan and its use as a pharmaceutical excipient, *Pharm. Res.*, 15: 1326-31, 1998.
 77. Illum L, Farraj NF, Davis SS. Chitosan as a novel delivery system for peptide drugs. *Pharm. Res.*, 11: 1186-89, 1994.
 78. Soane RJ, Frier M, Perkins AC, Jones NS, Davis SS, Illum L. Evaluation of the clearance characteristics of bioadhesive systems in humans. *Int. J. Pharm.*, 178: 55-65, 1999.
 79. Aspden TJ, Illum L, Skaugrud O. Chitosan as a nasal delivery system: Evaluation of insulin absorption enhancement and effect on nasal membrane integrity using rat models. *Eur. J. Pharmaceut. Sci.*, 4: 23-31, 1996.
 80. Dodane V, Vilivalam VD. Pharmaceutical applications of chitosan. *Pharmaceut. Sci. Technol. Today*, 1: 246-253, 1998.
 81. Fernandez-Urrusuno, R. Calvo P, Remunan-Lopez C, Vila-Jato JL, Alonso MJ. Enhancement of nasal absorption of insulin using chitosan nanoparticles. *Pharmaceut. Res.*, 16:1576-81, 1999.
 82. Illum L, Watts P, Fisher AN, Jabbal GI, Davis SS. Novel chitosan based delivery systems for the nasal administration of a LHRH-analogue. *STP Pharma. Sci.*, 10: 89-94, 2000.
 83. Janes K A, Calvo P, Alonso M J. Polysaccharide colloidal particles as delivery systems for macromolecules. *Adv. Drug Del. Rev.*, 47:83-97, 2001.
 84. Van der Lubben, IM et al. Chitosan and its derivatives in mucosal drug and vaccine delivery. *Eur. J. Pharmaceut. Sci.*, 14, 201-207, 2001.
 85. Dyer AM, Hinchcliffe M, Watts P, Castile J, Jabbal-Gill I, Nankervis R, Smith A, Illum L. Nasal delivery of insulin using novel chitosan based formulations: A comparative study in two animal models between simple chitosan formulations and chitosan nanoparticles. *Pharmaceut. Res.*, 19:998-1008, 2002.
 86. Hinchcliffe, M, Jabbal-Gill, I, Smith, A. Effect of chitosan on the intranasal absorption of salmon calcitonin in sheep. *J. Pharm. Pharmacol.*, 57, 681-687, 2005.
 87. Artursson, P. et al., Effect of chitosan on the permeability of monolayers of intestinal epithelial cells (Caco-2). *Pharm. Res.*, 11, 1358-61, 1994.

88. Schipper NGM, Romeijn SG, Verhoef JC, Merkus FWHM. Nasal insulin delivery with dimethyl-beta-cyclodextrin as an absorption enhancer in rabbits: powder more effective than liquid formulations. *Pharm. Res.*, 10(5):682-86, 1993.
89. Frauman AG, Jerums G, Louis WJ. Effects of intranasal insulin in non-obese type II diabetics. *Diabetes Research and Clinical Practice*, 3:197-202, 1987.
90. Deurloo MJ, Hermens WA, Romeyn SG, Verhoef JC, Merkus FW. Absorption enhancement of intranasally administered insulin by sodium taurodihydrofusidate (STDHF) in rabbits and rats. *Pharm. Res.*, 6:853-856, 1989.
91. Johansson F, Hjertberg E, Eirefelt S, Tronde A, Hultkvist Bengtsson U. Mechanisms for absorption enhancement of inhaled insulin by sodium taurocholate. *Eur. J. Pharm. Sci.*, 17, 63-71, 2002.
92. Li Y, Shao Z, Mitra AK. Dissociation of insulin oligomers by bile salt micelles and its effect on alpha-chymotrypsin-mediated proteolytic degradation. *Pharm. Res.*, 7:864 - 869, 1992.
93. Maggio ET. Alkylsaccharides: circumventing oxidative damage to biotherapeutics caused by polyoxyethylene-based surfactants. *Ther. Deliv.* 4(5):567-72, 2013.
94. Maggio ET. Polysorbates, peroxides, protein aggregation, and immunogenicity a growing concern. *J. Excipients and Food Chem.* 3 (2):45-53, 2012.
95. Fernández-Urrusuno R, Calvo P, Remuñán-López C, Vila-Jato JL, Alonso MJ. Enhancement of nasal absorption of insulin using chitosan nanoparticles. *Pharm. Res.* 16(10):1576-81, 1999.